OMC FILE CORY

SECURITY CLASSIFICATION OF THIS PAGE

REPORT DOCUMENTATION PAGE					Form Approved OMB No. 0704-0188
ELECTE NA 16 RESTRICTIVE MARKINGS					
ALIO O A 1000 STRIBUTION / AVAILABILITY OF REPORT					
AD-A197 704	Distribution unlimited				
4. PERFORMING ORGANIZATION REPORT NUMBER(S)		5. MONITORING ORGANIZATION REPORT NUMBER(S)			
Boston University ·		· NA			
6a NAME OF PERFORMING ORGANIZATION 6b OFFICE SYMBOL (If applicable)		7a. NAME OF MONITORING ORGANIZATION			
Boston University NA		Office of Naval Research			
6c. ADDRESS (City, State, and ZIP Code)	7b. ADDRESS (City, State, and ZIP Code)				
Chemistry Department Boston University	800 N. Quincy St. Arlington, VA 22217-5000				
590 Commonwealth Ave., Boston, MA 02215					
Ba. NAME OF FUNDING/SPONSORING ORGANIZATION	9. PROCUREMENT INSTRUMENT IDENTIFICATION NUMBER				
Office of Naval Research 8c. ADDRESS (City, State, and ZIP Code)	N00014-86-K-0217 10 SOURCE OF FUNDING NUMBERS				
800 N. Quincy St.		PROGRAM PROJECT TASK WORK UNIT			
Arlington, VA 22217-5000		ELEMENT NO.	NO RR04106	NO. 441c0	ACCESSION NO.
11. TITLE (Include Security Classification)					
(U) Characterization of Marine Bioadhesive Proteins					
12 PERSONAL AUTHOR(S) Laursen, Richard A.					
13a TYPE OF REPORT 13b TIME COVERED 14. DATE OF REPORT (Year, Month, Day) 15 PAGE COUNT					
Annual FROM //1/8/ TO 6/30/8 1988 June 10 3					
TO SOLVEDWICK AND MOVEMENT AND					
17 COSATI CODES FIELD GROUP SUB-GROUP		Continue on reverse if necessary and identify by block number) roteins, Mussel, Amino Acid			
TIELD GROOF SOB-GROOF		ene sequence, cDNA			
(mgm)					
19 ABSTRACT (Continue on reverse if necessary and identify by block number)					
The objective of this research is to elucidate the amino acid sequences, via					
gene sequencing, of the adhesive proteins from several species of mussel and					
of other organisms, with the aim of understanding how these organisms attach					
themselves to wet surfaces. During the past year, we have cloned and sequenced fragments of the adhesive protein genes from three species of					
mussel. Two classes of protein are now apparent: they are similar in their					
content and location of lysine and tyrosine (or DOPA) residues, but different					
in repeat length and content of other amino acids.					
20 DISTRIBUTION / AVAILABILITY OF ABSTR	21 ABSTRACT SECURITY CLASSIFICATION				
UNCLASSIFIED/UNLIMITED SAME AS RPT DTIC USERS 22. NAME OF RESPONSIBLE INDIVIDUAL Dr. E. Smell, M. Marron or M. Haygood		(U) 226 TELEPHONE (I (202) 696-4	nclude Area Code) 22c OF	
DD Form 1473, JUN 86 Previous editions are obsolete SECURITY CLASSIFICATION OF THIS PAGE					
S/N 0102-LF-014-6603 DISTRIBUTION STATEMENT A					

Approved for public release; Distribution Unlimited

PROGRESS REPORT ON CONTRACT NOO014-86-K-0217

PRINCIPAL INVESTIGATOR: Richard A. Laursen

CONTRACTOR: Boston University

CONTRACT TITLE: Characterization of Marine Bioadhesive Proteins

START DATE: 1 April 1986

RESEARCH OBJECTIVE: The primary initial objective has been to clone and sequence adhesive protein genes for several species of mussel with the aim of understanding what common (if any) structural features give these proteins their adhesive properties. It is hoped that this knowledge will lead to the development of adhesives that will have medical and other applications.

PROGRESS (YEAR 2): During the first year and continuing into the second, the focus of our work was isolating and sequencing several cDNA clones of fragments of the adhesive protein gene. This work showed that adhesive protein of M. edulis is primarily repeats of the decapeptide

xx1-Lys-xx2-xx3-Tyr-Pro-Pro-Thr-Tyr-Lys

where xx1 is usually Pro, Ser or Ala; xx2 is Pro, Ser, Leu, Ile or Lys; and xx3 is Thr or Ser. Using our original methods, however, we have not been able to obtain a clone or set of overlapping clones that encode for the entire protein. It appears that recombination, due to the repetitive nature of the gene, is occurring during cloning. Recently we have tried to overcome this problem we have fractionated our cDNA library, selected a fraction (3.3 kbp) large enough to code for the entire protein and are carrying out the subsequent cloning steps in recombinant-minus host strains. We have also isolated M. edulis genomic DNA and are currently screening the genomic library.

In year 2 we have also obtained sequence data from two other species of mussel, Mytilus californianus and Geukensia demissus, and we will soon have data from Modiolus modiolus. Cloning of M. californianus and of M. modiolus genes was carried out as for M. edulis by construction of a Agt10 cDNA library and screening with probes from M. edulis. The sequence of a clone from M. californianus was very simililar to that of M. edulis, except for the occurence of Arg (50% of the time) at position xx1 and about a 50% occurence of Ser and Ala at position xx7.

Cloning the G. demissa gene: A \(\) \(\) \(\) \(\) this species, but screening with \(\) \\(\) \

The λ gtll library was packaged into phage and used to infect *E. coli* Y1090 host cells, and β -galactosidase fusion products were detected as colorless plaques among blue plaques in nonrecombinants. Colorless plaques were replated and screened with adhesive protein antibodies and an alkaline phosphatase linked secondary antibody, which gave a blue color with the substrate BCIP (5-bromo-4-chlor-3-indolylphosphate-p-toluidide). Positive clones were then sequenced in the usual manner. The *G. demissa* protein is significalntly different in that it contains repeats of from 11 to 13 amino acids, e.g.,

Gly-Lys-Pro-Thr-Thr-Tyr-Asp-Ala-Gly-Tyr-Lys-Gly-Gln-Lys-Gln-Thr-Gly-Tyr-Asp-Thr-Gly-Tyr-Lys-,

and contains large amounts of glycine and glutamine, but little proline. Genetic material for this species was obtained by immunoscreening a λ gt11 cDNA library, because, in contrast with the other species, we had no protein sequence data to guide the synthesis of oligonucleotide probes.

The recombination problem: Northern blot experiments in which a \$32p-labeled oligonucleotide probe is allowed to hybridize with mRNA have consistently shown, for all species, that the mRNA we have isolated from mussel phenol glands is long enough (3.0-4.0 kbp) to code for an adhesive protein with a molecular weight of up to 130,000. However screening of cDNA libraries shows not only many fewer clones than we might expect, but also much smaller, typically less than 500 base pairs, than the 3500 bp needed to code for the entire protein. This suggests that during the cloning process large amounts of information are being recombined out, even though we are using RecA- host strains. Furthermore, the fragments we have sequenced, and also those sequenced at Genex Corp. (unpublished), do not overlap, despite the fact that we have enough data to account for more than the entire protein. So the situation may be even worse than loss of information, there may also be scrambling.

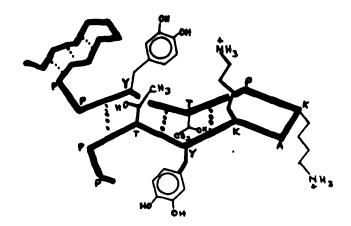
Other workers have also encountered difficulty in cloning repetitious DNA sequences in certain E. coli strains. To overcome this problem, we are now beginning to carry out cloning operations in recommendated in the commentation of E and E and E and E are the commentation of E are the commentation of E and E are the commentation of E

A conformational model for the Mytilus protein: Because of the invariability of Tyr and Lys residues and the patterns of posttranslational modification of Tyr and Pro residues, we believe that the adhesive proteins probably have some sort of regular, as opposed to a "random coil", structure. Given the large amount of proline, a structure with turns or loops seems more likely than a regular helical or sheet structure. Given the propensity for Tyr and Thr residues to occur in β -sheets, for Pro-Pro sequences not to be found in β -turns, but to cause a 90° bend in the peptide backbone, we have postulated the following β -sheet- β -turn model to serve as a working hypothesis for planned spectroscopic studies:

DTIC Special Special COPY (NSPECTED)

Page 2

Codes



This model, though speculative, has some attractive features. It puts all the polar groups on the faces of the β -sheet loop, where they could interact with surfaces. In addition the Tyr and Lys residues are on both faces in pairs, in a symmetrical arrangement, where they might pair up with corresponding pairs in another chain to form interchain crosslinks. the major failing of this model is that one cannot make a similar model for the Geukensia protein, which contains little proline and has a less regular repeat structure. Of course Geukensia could have a completely different structure, but one would think, given the relatively constant placement of the critical Tyr and Lys residues, that there might be some conformational similarities. The answer to this dilemma can be answered only by experiment.

WORK PLAN (YEAR 3) In year three we plan to concentrate on obtaining the entire sequence of an adhesive protein either by sequencing genomic DNA or through the use of the rec-cloning strains mentioned above. Even if that fails, we now have or soon will have sufficient sequence data to begin analyzing the problem of what gives this class of proteins their adhesive character. During the next year we will focus on obtaining more sequence data on the G. demissa protein, because it is so different from the other species, and on getting sequence data from M. modiolus.

We plan also to begin conformational and modeling studies, using high resolution NMR techniques, on the proteins or peptide models, since it seems likely to us that these proteins have some sort of regular structure. If time permits, we hope to characterize the crosslink, which is presumed to occur between lysine and DOPA residues in these proteins, using chemical and mass spectrometric methods.